



## Effect of Plant Growth Regulators Effect on Grape Cutting (*Vitis vinifera* L.) cv. Flame Seedless

Gurdeep Singh and Navdeep Singh

Department of Horticulture

Guru Kashi University, Talwandi Sabo, Bathinda-151302, Punjab, India

### ABSTRACT

The present investigation was conducted at Guru Kashi University Research Farm during 2022-2023, Punjab on evaluating the effect of plant growth regulator on grape cv. Flame seedless cutting. These cutting was treated with different concentration of indole-3-butyric acid ( $T_1$ : 1000,  $T_2$ : 1500 and  $T_3$ : 2000 ppm) and gibberellic acid ( $T_4$ : 50,  $T_5$ : 100 and  $T_6$ : 150 ppm) along with control ( $T_0$ ) using prolonged dipping method. The result revealed that  $T_6$ : gibberellic acid @150 ppm (9.33) showed minimum numbers of days taken for first emergence of nodes whereas,  $T_2$ : indole-3-butyric acid @ 1500 ppm (8.67) showed minimum days of the first roots emergence. The maximum survival percentage was recorded in the cutting treated with  $T_3$ : indole-3-butyric acid 2000 ppm (80.00%). Hence,  $T_2$ : indole-3-butyric acid @ 1500 ppm was showed good vegetative and root growth in selected root cutting of grape (*Vitis vinifera* L.) cv. Flame Seedless.

**Key Word:** Cutting, Grape, Growth regulators, Root growth, Survival.

### INTRODUCTION

Grape (*Vitis vinifera* L.), belongs to family Vitaceae is important commercial fruit crops with extraordinary taste and flavor cultivated in temperate and tropical regions of World (Gowda *et al*, 2008; Nowshehri *et al*, 2015). The major grape-growing states in India, are Maharashtra (70.67%), Karnataka (24.49%), Tamil Nadu (1.43%), Andhra Pradesh (1.34%), Madhya Pradesh (1.02%), and Mizoram (0.50%), collectively contributes to nearly (99%) of the total grape production (NHB, 2019). Out of total production of grapes (74.5%) is used as table purpose, (23.5%) used as resin, (15%) used as wine and (0.5%) used for juices according to Adsule *et al* (2012). It possesses high nutrient content (10.2%) carbohydrates, (0.8%) proteins, (0.1%) minerals and (85.5%) water, fiber (1.4 g), vitamin C (27%), vitamin K (28%), thiamine (7%), riboflavin (6%), vitamin B6 (6%), potassium (8%), copper (10%), manganese (5%) and flavonoids (flavonoids, anthocyanins, and flavonols) as an important source of antioxidants

(Andjelkovic *et al*, 2013; Somkuwar *et al*, 2018). This nutrient content used in curing jaundice, asthma, joints pains, piles, diabetes, cancer and heart diseases (Kanagarla *et al*, 2013; Dohadwala *et al*, 2009).

Flame seedless cultivar is heavy bearing, crimson red colour, sweet flavour berries and most tolerant to pre monsoon rains as well as less susceptible to diseases according to Chanana *et al* (2008). The hard wood cutting is most common way of propagation of grape. In this method the treatment of cuttings with plant growth regulators play an important role in regeneration. Many scientists had already reported the role of these plant growth regulators in stem elongation, apical dominance, root initiation, increasing the root number and length (Ling *et al*, 2015) So, The present study was undertaken with objectives to evaluate the effect of Gibberellic Acid ( $GA_3$ ) and Indole Butyric Acid (IBA) on the survival percentage and vegetative growth of grapes cutting (*Vitis vinifera* L.) cv. Flame seedless.

## MATERIALS AND METHODS

The experiment was conducted at agriculture field of Guru Kashi University Research Farm, Talwandi sabo, Punjab in 2022-2023. It is located at latitude 29°59'0" N and longitude 75°5'0" East, has semiarid climate with wide variations of summer and winter temperatures. 20-25cm long grape (*Vitis vinifera* L. cv. Flame seedless) healthy, well nourished, and mature wood was used for trial. Basal 3-4 cm portion of hard wood cuttings were dipped for 24 h in indole-3-butyric acid ( $T_1$ : 1000,  $T_2$ : 1500 and  $T_3$ : 2000 ppm) and gibberellic acid ( $T_4$ : 50,  $T_5$ : 100 and  $T_6$ : 150 ppm) solution along with water as control ( $T_0$ ). It was allowed to dry for 15 minutes and then transplanted in field. The research trail was laid out in Randomized Block Designs (RBD). The treatments in experiment were replicated three times and each plot consisted of 30 cuttings. Further observations was recorded from parameters i.e. first emergence of node, first emergence of roots, survival percentage, average number of leaves, average number of roots from selected plants. All data from the experimental field were analyzed separately for each experiment for different growth characters and yield attributes with the help of OPSTAT (Statistical Software Package for Agricultural Research Workers) (Sheoran *et al*, 1998). The critical difference at 5% level of implication was calculated to equate the mean different treatments.

## RESULTS AND DISCUSSION

The result revealed that in  $T_6$ : GA<sub>3</sub>@150 ppm (9.33) least numbers of days taken for first emergence of nodes followed by  $T_5$ : GA<sub>3</sub>@100 ppm (11.33) and  $T_4$ : GA<sub>3</sub>@ 50 ppm (14.33) and  $T_0$ : control (20.67) maximum number of days was taken for first emergence of nodes. The result also revealed that  $T_2$ : IBA @ 1500 ppm (8.67) least numbers of days taken for first emergence of roots followed by  $T_3$ : IBA @ 2000 ppm (11.67),  $T_1$ : IBA @1000 ppm (14.33),  $T_6$ : GA<sub>3</sub>@150 ppm (19.33),  $T_4$ : GA<sub>3</sub>@ 50 ppm (18.67) and maximum number of days was taken for first emergence of roots in  $T_0$ : control (21.33). The maximum survival percentage was recorded in the cutting treated with  $T_3$  IBA @ 2000 ppm (80.00%) followed by  $T_2$  IBA @ 1500 ppm (78.33%),  $T_1$ : IBA @1000 ppm

(73.33%),  $T_6$ : GA<sub>3</sub>@150 ppm (72.33%),  $T_5$ : GA<sub>3</sub>@ 100 ppm (71.33%) and  $T_4$ : GA<sub>3</sub>@ 50 ppm (69.00%) and whereas, the minimum survival percentage was recorded in  $T_0$ : Control (58.33%). However, Patil *et al* (2000) reported the cuttings for 6 hours either in IBA (100 ppm) recorded maximum survival percentage (86.33 and 76.00) in the cutltivars Tas-A-Ganesh and Kismish Chorny.

The maximum average number of leaves at 60 and 90 DAP was observed in  $T_2$ : IBA @ 1500 ppm (20.67 & 25.33), followed by  $T_1$ : IBA @1000 ppm (13.67 & 22.33),  $T_3$ : IBA @ 2000 ppm (10.67 & 18.33),  $T_5$ : GA<sub>3</sub>@ 100 ppm (10.00 & 15.67),  $T_6$ : GA<sub>3</sub>@150 ppm (9.33 & 13.67),  $T_4$ : GA<sub>3</sub>@50 ppm (9.33 & 12.67), Whereas, the minimum average number of leaves was recorded in  $T_0$ : Control (8.67 & 9. 67) at 60 DAP and at 90 DAP. Many researchers work on the effect of plant growth regulators on the grapes cutting growth. Chalapathi *et al* (2001) who reported superior result in shoot length, number of branches, number of leaves and root length, survival percentage and sprouting percentage after cuttings treated with IBA.

The maximum average number of roots was recorded in the cutting treated with  $T_2$ : IBA @ 1500 ppm (31.67), followed by  $T_3$ : IBA @ 2000 ppm (30.00),  $T_1$ : IBA @1000 ppm (27.33),  $T_6$ : GA<sub>3</sub>@150 ppm (23.33),  $T_5$ : GA<sub>3</sub>@ 100 ppm (22.33), and  $T_4$ : GA<sub>3</sub>@ 50 ppm (20.33). Whereas, the minimum average number of roots was recorded in  $T_0$ : Control (9. 67). Patil *et al* (2001) also reported the better survival percentage and higher number of primary roots with IBA treatments. Similarly, Song *et al* (2001) reported good rooting when the base of the cuttings was soaked in a solution of 150 ppm IBA for 24 hours. Rao (2004) reported IBA, 2000 ppm for hardwood cuttings was good for highest percentage of rooting and number of roots and longest root length per cutting in Dogridge and 1613C rootstocks. The maximum numbers of roots were obtained in grapes (*V. vinifera*) with 4000 mg/l IBA. The study showed the significant positive effect of both IBA and GA<sub>3</sub> plant growth regulator on growth and survival of grapes cutting cv. Flame seedless. The IBA @ 1500 ppm has shown good

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**Table I. Effect of plant growth regulators on total numbers of days taken for first emergence of node and roots (Days), survival percentage (%), average number of leaves / cutting (No.) and average number of roots/ cutting (No.) on grapes cutting (*Vitis vinifera* L.) cv. Flame Seedless.**

| Treatment | Number of days taken for first emergence of node (Days) | Numbers of days taken for first emergence of roots (Days) | Survival percentage (%) | Average Number of leaves / cutting (No.) |        | Average Number of roots/ cutting (No.) |
|-----------|---|---|-------------------------|--|--------|--|
|           |   |   |                         | 60 DAP                                   | 90 DAP |  |
| T0        | 20.67   | 21.33   | 58.33                   | 8.67                                     | 9.67   | 15.33                                  |
| T1        | 19  | 14.33   | 73.33                   | 13.67                                    | 22.33  | 27.33                                  |
| T2        | 16.33   | 8.67  | 78.33                   | 20.67                                    | 25.33  | 31.67                                  |
| T3        | 18.33   | 11.67   | 80                      | 10.67                                    | 18.33  | 30                                     |
| T4        | 14.33   | 18.67   | 69                      | 9.33                                     | 12.67  | 20.33                                  |
| T5        | 11.33   | 16.67   | 71.33                   | 10                                       | 15.67  | 22.33                                  |
| T6        | 9.33  | 19.33   | 72.33                   | 9.33                                     | 13.67  | 24.33                                  |
| CV%       | 6.82%   | 3.44%   | 3.77%                   | 9.59%                                    | 4.34%  | 2.95%                                  |
| SE±       | 0.62  | 0.35  | 1.61                    | 0.52                                     | 0.32   | 0.41                                   |

vegetative growth, less number of days taken for first emergence, average number leaves and root growth in root cutting of grape (*Vitis vinifera* L.) cv. Flame seedless.

### CONCLUSION

It was concluded that the maximum data observed of cutting survival percentage in IBA 2000 ppm treatments. Moreover both plant growth regulators IBA and GA<sub>3</sub> showed significant effect on the growth and development of grapes as compared to control.

### REFERENCES

- Adsule P G, Kumar S A, Anuradha U, Indu S, Satisha J, Upadhyay A K and Yadav D S (2012). Grape Research in India -A Review. *Progr Hort* **44**(2): 180-193.
- Andjelkovic M, Radovanovic B, Radovanovic A and Andjelkovic A M (2013). Changes in polyphenolic content and antioxidant activity of grapes cv. vranac during ripening. *S African J Enol Vitic* **34**(2):147-155.
- Anonymous (2019). *Area under grape cultivation in India*. National Horticulture Board 68-75.
- Chalapathi M V, Thimmegowda N D, Kumar S, Gangadhar G, Rao E and Mallikarjun K (2001). Influence of length of cutting and growth regulators on vegetative propagation of Stevia (*Stevia rebaudiana* Bert.). *Crop Res* **21**: 53-56.
- Chanana Y R and Gill M I S (2008). High quality grapes can be produced in Punjab. *Acta Hort* **785**: 85-86.
- Dohadwala M M and Vita J A (2009). Grapes and cardiovascular disease. *J Nutr* **139** (9):1788S-93S.
- Galavi M, Karimian M A, Mousavi S R (2013). Effects of different auxin (IBA) concentrations and planting-beds on rooting grape cuttings (*Vitis vinifera*). *Annu Res Rev Biolo* **3**(4):517-523.
- Gowda V N, Keshava S A and Shyamamma S (2008). Growth, yield and quality of Bangalore Blue grapes as influenced by foliar applied polyfeed and multi-K. Proceedings of the International Symposium on Grape Production and Processing. *Acta Hort* **785**: 207-212.
- Kanagarla N S S A, Kuppast I J, Veerashekar T and Reddy L R (2013). A review on benefits and uses of *Vitis vinifera* (Grape). *Res and Review in Biosci* **7**(5): 175-180.
- Ling G, Murphy A S, Peer W A, Gan L, Yi Li Y and Cheng Z (2015). Physiological and Molecular Regulation of Adventitious Root Formation. *Crit Rev Pl Sci* **34** (5): 506-521.
- Nowshehri J, Bhat Z and Shah M (2015). Blessings in disguise: Bio-functional benefits of grape seed extracts. *Food Res Int* **77**(3): 333-348.

- Patil V N, Chauhan P S, Panchabhai D M, Shivankar R S and Tannirawar A V (2000). Effect of growth regulators on rooting of hardwood cuttings of some commercial grape varieties. *J Soil Crop* **10** (2): 295-297.
- Patil V N, Chauhan P S, Shivankar R S, Vilhekar S H and Waghmare V S (2001). Effect of plant growth regulators on survival and vegetative growth of grapevine cuttings. *Agri Sci Dig* **21**(2): 97-99.
- Rao K K (2004). *Studies on the propagation of grape rootstocks through hardwood and soft wood cuttings*. MSc. thesis. Rajendranagar, Hyderabad, pp. 1-97
- Sheoran O P, Tonk D S, Kaushik L S, Hasija R C and Pannu R S (1998). Statistical Software Package for Agricultural Research Workers. Recent Advances in information theory, Statistics & Computer Applications by D.S. Hooda & R.C. Hasija Department of Mathematics Statistics, CCS HAU, Hisar, pp. 139-143.
- Somkuwar R G, Bhange M A, Oulkar D P, Sharma A K and Ahammed S T P (2018). Estimation of polyphenols by using HPLC–DAD in red and white wine grape varieties grown under tropical conditions of India. *J Food Sci Technol* **55** (12): 4994-5002.
- Song Y G, Lu W P, Wang J, Shen Y J, Wu Z and Liu W D (2001). Study on promoting the rooting ability of hardwood cuttings of Amurien grape varieties. *China Fruits* 1: 4-7.

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